

afforded 44 mg lupeyl acetate and 11 mg caryophyllen-1,10-epoxide, while TLC of fraction 3 (CH_2Cl_2 - C_6H_6 - Et_2O , 1:1:1) gave a mixture of 2-4 and the known compounds described in the text. Separation by HPLC (RP 8, MeOH - H_2O , 3:2, flow rate ca 3 ml/min and 300 bar) gave a mixture of 4 and onoseriolide (I, R_f 10.1 min), a mixture of the latter, 2 and 3 (II, R_f 11.4 min) and a mixture of 2, 3, alantolactone and onoseriolide (III, R_f 12.2 min). TLC of I (SiO_2 , AgNO_3 -coated, Et_2O -petrol, 2:3, 2 developments) gave 4 mg of the latter and 2.5 mg 4 (R_f 0.3). TLC of II (SiO_2 , AgNO_3 -coated, Et_2O -petrol, 2:3, 2 developments) gave 2.5 mg onoseriolide, 2.1 mg 3 and 1 mg 2, while TLC of III (SiO_2 , AgNO_3 -coated, Et_2O -petrol, 2:3, 2 developments) afforded 2 mg onoseriolide, 3 mg alantolactone and 2.5 mg 3.

1,2,4,15-Tetradehydro-4,5-dihydrosteuractinolide (4) Colourless oil, IR $\nu_{\text{CCl}_4} \text{ cm}^{-1}$ 1770 (γ -lactone), MS m/z (rel int) 230 146 $[\text{M}]^+$ (5) (calc for $\text{C}_{15}\text{H}_{18}\text{O}_2$ 230 146), 215 $[\text{M} - \text{Me}]^+$ (2.5), 173 $[\text{215} - \text{C}_3\text{H}_6]^+$ (100), $^1\text{H NMR}$ (CDCl_3 , 400 MHz, TMS as internal standard) δ 5.39 *dt* (H-1), 5.52 *dt* (H-2), 2.89 *br d* and 2.74 *br dd* (H-3), 2.10 *br d* (H-5), 2.10 *br dd* (H-6), 1.93 *ddd* (H-6'), 3.33 *m* (H-7), 4.80 *ddd* (H-8), 2.15 *br d* (H-9), 1.33 *dd* (H-9'), 6.32 *d* and 5.54 *d* (H-13), 0.80 *s* (H-14), 4.93 and 4.71 *br s* (H-15), $[J \text{ (Hz)}]$ 1, 2 = 10, 1, 3 = 2, 2, 3 = 3.5, 5, 6' = 6, 6' = 6', 7 = 12, 5, 6 = 6, 7, 8 = 7, 3, 17 = 3.5, 7, 13' = 3, 8, 9 ~ 7, 8, 9' = 11, 9, 9' = 13].

Callurin [7] $^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 5.75 *dd* (H-1), 4.93 *dd* (H-2c), 4.89 *dd* (H-2t), 4.89 *dq* (H-3), 4.64 *br s* (H-3'), 1.67 *br d* (H-5), 2.06 *m* (H-6), 2.30 *m* (H-7), 4.60 *ddd* (H-8), 1.90 *dd* (H-9), 1.40 *dd* (H-9'), 2.56 *dq* (H-11), 1.20 *d* (H-13), 1.00 *s* (H-14), 1.72 *br s* (H-15), $[J \text{ (Hz)}]$ 1, 2c = 10, 1, 2t = 17, 2c, 2t = 1, 3, 5

= 3, 15 = 1.5, 5, 6 = 12, 7, 8 = 8, 7, 11 = 12, 8, 9 = 6, 8, 9' = 11, 9, 9' = 14, 11, 13 = 7].

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FURTHER CADINENE DERIVATIVES FROM *HETEROOTHECA LATIFOLIA*

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Key Word Index—*Heterotheca latifolia*, Compositae, sesquiterpenes, cadinene derivatives

Abstract—A reinvestigation of the aerial parts of *Heterotheca latifolia* afforded four new cadinene derivatives

So far the chemical investigations of *Heterotheca* species have shown that cadinene derivatives are characteristic for this genus [1, 2]. A reinvestigation of the aerial parts of *H. latifolia* Buckley afforded in addition to compounds isolated previously [1] the cadinene derivatives 1-4 which were isolated as their methyl esters (1a-4a). The structures of 1a and 2a could be deduced from the $^1\text{H NMR}$ spectral data (Table 1) which were close to those of the corresponding esters of 1a [1]. Also the $^{13}\text{C NMR}$ spectrum (see Experimental) supported the structure of 1a which finally was established by saponification of the cor-

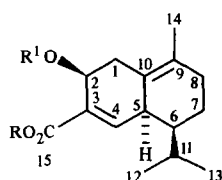
responding acetate [1]. After addition of diazomethane, a methyl ester was obtained which was identical with 1a. The structures of 3a and 4a also could be deduced from the $^1\text{H NMR}$ spectra (Table 1). The presence of hydroperoxides followed from the low-field broadened singlets at δ 7.45 and 7.28, respectively, while several signals were close to those of methyl-13-hydroxy- δ -cadinen-15-oate [1]. However, in the spectrum of 3a the Δ^9 bond was replaced by a 9,14-double bond, which followed from the typical signals of exocyclic olefinic protons. Their chemical shifts already indicated that the hydroperoxy group

Table 1 ^1H NMR spectral data of **1a–4a** (400 MHz, CDCl_3 , TMS as int standard)

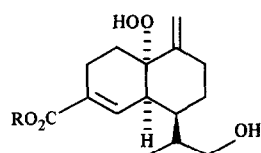
	1a	C_6D_6	2a	3a	4a
H-1	2.99 <i>dd</i>	2.99 <i>dd</i>	3.09 <i>dd</i>	2.14 <i>m</i>	} 5.86 <i>br dd</i>
H-1'	1.95 <i>m</i>	1.83 <i>m</i>	1.80 <i>br d</i>	1.70 <i>br d</i>	
H-2	} 4.65 <i>br s</i>	} 4.79 <i>br s</i>	} 4.24 <i>br s</i>	} 2.37 <i>m</i>	} 3.10 <i>dddd</i>
H-2'					
H-4	7.12 <i>d</i>	7.21 <i>d</i>	7.15 <i>d</i>	7.08 <i>ddd</i>	7.25 <i>ddd</i>
H-5	2.66 <i>br d</i>	2.50 <i>br d</i>	2.63 <i>br d</i>	2.98 <i>br dd</i>	3.30 <i>dddd</i>
H-6	1.25 <i>m</i>	1.16 <i>dddd</i>	1.28 <i>m</i>	1.28 <i>m</i>	1.3 <i>m</i>
H-7		1.50 <i>br d</i>		1.73 <i>m</i>	1.67 <i>m</i>
H-7'		1.09 <i>dddd</i>		1.29 <i>m</i>	1.48 <i>m</i>
H-8	} 2.03 <i>m</i>	} 1.89 <i>br dd</i>	} 2.05 <i>br dd</i>	} 2.47 <i>ddd</i>	} 2.22 <i>m</i>
H-8'					
H-11	2.12 <i>dqq</i>	1.98 <i>dqq</i>	2.13 <i>dqq</i>	2.14 <i>dqq</i>	2.22 <i>dqq</i>
H-12	0.99 <i>d</i>	0.84 <i>d</i>	0.97 <i>d</i>	1.01 <i>d</i>	1.06 <i>d</i>
H-13	} 0.82 <i>d</i>	} 0.69 <i>d</i>	} 0.81 <i>d</i>	} 3.78 <i>dd</i>	} 3.82 <i>dd</i>
H-13'					
H-14	1.75 <i>br s</i>	1.75 <i>br s</i>	1.71 <i>br s</i>	} 5.15 <i>dd</i> 4.97 <i>dd</i>	} 1.51 <i>s</i>
OMe	3.78 <i>s</i>	3.59 <i>s</i>	3.76 <i>s</i> 3.42 <i>s</i>	3.74 <i>s</i>	3.84 <i>s</i>
OOH	—	—	—	7.45 <i>br s</i>	7.28 <i>br s</i>

J (Hz) compounds **1a** and **2a** 1, 1' = 15, 1, 2 = 2.5, 1', 2 ~ 2, 4, 5 = 2, 5, 6 = 8, 7, 7' = 12, 7, 8 = 12, 7', 8 = 4.5, 8, 8' = 15, 11, 12 = 11, 12 = 11, 13 = 7, compound **3a** 1, 1' = 14, 2, 4 = 2', 4 = 1.5, 4, 5 = 4, 5, 6 = 10, 7, 8 = 7', 8 = 4, 8, 8' = 14.5, 8, 14 = 14, 14' ~ 1.5, 11, 13 = 5, 11, 13' = 7, 13, 13' = 10.5, compound **4a** 1, 2 = 1, 2' = 3.5, 2, 2' = 2.2, 3, 4 = 1.5, 2, 5 = 7, 2', 4 = 1, 2', 5 = 7, 4, 5 = 4, 5, 6 ~ 10, 7, 8 = 7', 8 = 3, 8, 8' = 14, 11, 13 = 6, 11, 13' = 8, 13, 13' = 10

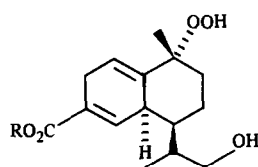
most likely was at C-10. Though only one of the H-8 signals was not overlapped, the clear couplings of this one showed that H-8' was coupled with H-14. Irradiation of the signal at δ 2.22 collapsed the methyl doublet to a singlet and the H-13 double doublets to doublets. The oxygen function at C-10 is most likely α as the H-6 signal is shifted downfield when compared with that in the cadin-



	1	1a	2	2a
R	H	Me	H	Me
R ¹	H	H	Me	Me



3 R=H
3a R=Me



4 R=H
4a R=Me

3-enes. The presence of a 1(10), 3-diene in the hydroperoxide **4a** clearly followed from the downfield shift of the signals of H-2 and H-5. A large coupling (~ 7 Hz) between H-2 and H-5 further supported the presence of such a diene. As in this case the signal of the olefinic methyl at C-9 was replaced by a singlet at δ 1.51, the hydroperoxy group was at C-9. Inspection of a model indicated that an α -orientation was more likely as the attack of oxygen from the α side of methyl-13-hydroxy- δ -cadinen-15-oate seems to be favoured for steric reasons. Compounds **3** and **4** are probably formed by reaction of 13-hydroxy- δ -cadinen-15-oic acid [1] with oxygen. This investigation again showed the taxonomic importance of cadinene derivatives for distinguishing the genus *Heterotheca* from other related genera.

EXPERIMENTAL

The air-dried aerial parts (210 g) (voucher RMK 9311, collected in DE, U.S.A.) were worked up in the usual fashion [3] and one-eighth of the CC fraction with Et_2O (280 mg) was esterified with CH_2N_2 . TLC (Et_2O -petrol, 1:1) gave 85 mg of the esters of **1a** (2-*O*-isobutyrate, 2-*O*-isovalerate, 2-methylbutyrate), 52 mg of the acetate of **1a**, a mixture of **1a** and **3a** as well as a mixture of **3a** and **4a**. Both mixtures were separated by TLC (Et_2O -petrol, 4:1). The less polar fraction afforded 10 mg **1a** (R_f 0.62) and 5 mg **3a** (R_f 0.53), and the more polar fraction gave 12 mg **4a** (R_f 0.45). One-sixth of the CC fraction with Et_2O -MeOH, 9:1 (250 mg) was esterified with CH_2N_2 . TLC (Et_2O -petrol, 7:3) gave a mixture of **1a** and **2a** as well as 60 mg **1a** and 50 mg methyl-13-

hydroxy- δ -cadinen-15-oate TLC of the mixture of **1a** and **2a** (Et₂O-petrol, 3:2) gave 70 mg **2a** (*R_f* 0.70). Known compounds were identified by comparison of the 400 MHz ¹H NMR spectra with those of authentic material and by co-TLC.

Methyl 2 β -hydroxy- δ -cadinen-15-oate (1a) Colourless oil, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹ 3560 (OH, hydrogen bonded), 1710, 1655 (C=CCO₂R), MS *m/z* (rel int) 264 [M]⁺ (24) (calc for C₁₆H₂₄O₃ 264.173), 246 [M-H₂O]⁺ (20), 203 [246-CHMe₂]⁺ (80), 187 [246-CO₂Me]⁺ (100), 176 [246-H₂C=CHCHMe₂, RDA]⁺ (30), ¹³C NMR (C₆D₆, C-1-C-15) 33.2 t, 65.4 d, 124.0 s, 144.4 d, 41.0 d, 44.0 d, 21.7 t, 34.6 t, 129.6 s, 133.0 s, 27.0 d, 19.1 q, 21.5 q, 15.7 q, 16.7 s, 51.3 q (OMe).

Preparation of 1a from the acetate To 23 mg β -acetoxy- δ -cadinen-15-oic acid in 2 ml MeOH, 0.5 ml 2 N KOH was added. After 2 hr the crude acid was esterified with CH₂N₂. TLC (Et₂O-petrol, 4:1) gave 16 mg **1a** (*R_f* 0.60) identical with the methyl ester obtained from the natural product (¹H NMR and co-TLC).

Methyl 2 β -methoxy- δ -cadinen-15-oate (2a) Colourless oil, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹ 1710, 1650 (C=CCO₂R), MS *m/z* (rel int) 278 [M]⁺ (11) (calc for C₁₇H₂₆O₃ 278.188), 260 [M-H₂O]⁺ (7), 246 [M-MeOH]⁺ (51), 235 [M-CHMe₂]⁺ (9), 203 [235-MeOH]⁺ (100), 187 [246-CO₂Me]⁺ (91), 176 [246-H₂C=CHCHMe₂, RDA]⁺ (45), 145 [187-C₃H₆]⁺ (76).

Methyl 13-hydroxy-10 α -peroxy-cadina-3,9(14)-dien-15-oate (3a) Colourless oil, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹ 3600 (OH), 1710 (C=CCO₂R), MS (CI, isobutane) *m/z* (rel int) 297 [M+1]⁺ (62) (calc for C₁₆H₂₄O₅+1), 279 [297-H₂O]⁺ (100), 263 [297-H₂O₂]⁺ (57), 247 [279-MeOH]⁺ (38), EI 262 [M-H₂O₂]⁺ (21), 231 [262-OMe]⁺ (32), 203 [231-CO]⁺ (42), 61 (100),

$$[\alpha]_{24}^{20} = \frac{578}{-10} \frac{546}{-19} \frac{436 \text{ nm}}{-63} \text{CHCl}_3, c = 0.3$$

Methyl 13-hydroxy-9 α -peroxy-cadina-1(10),3-dien-15-oate (4a) Colourless oil, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm⁻¹ 3600 (OH), 1720 (C=CCO₂R), MS (CI, isobutane) *m/z* (rel int) 297 [M+1]⁺ (10), (calc for C₁₆H₂₄O₅+1), 279 [297-H₂O]⁺ (21), 263 [297-H₂O₂]⁺ (14), 247 [279-MeOH]⁺ (8), 209 [297-C₅H₁₂O]⁺ (100), EI 262 [M-H₂O₂]⁺ (2.5), 230 [262-MeOH]⁺ (40), 61 (100).

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EUDESMANOLIDES FROM ARTEMISIA JUDAICA

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Key Word Index—*Artemisia judaica*, Compositae, sesquiterpenes, eudesmanolides

Abstract—The aerial parts of *Artemisia judaica* afforded in addition to known compounds a hydroperoxide corresponding to vulgarin as well as an isomer of the latter. The configurations of these lactones have been established by NOE difference spectroscopy.

The aerial parts of *Artemisia judaica* L. have been investigated previously [1-3], and we have studied now material collected in Egypt. In addition to tauremisin (= vulgarin, **1**) [4, 5] isolated previously from this species [1, 2], we obtained the isomer **3** and the hydroperoxide **2** and in addition ethyl cinnamate, α -pinene, chrysanthenone, camphor, piperitone, verbenol and the hydroperoxide **4** isolated so far only from *Artemisia inculta* [6]. The structure of **2** could be deduced from the spectral data of the product obtained by triphenyl phosphine-reduction which were identical with those of **1**. The spectral data of **3** (Table 1) were close to those of **1**. However, the chemical shifts of H-14 and H-15 differed characteristically. NOE

difference spectroscopy with both **1** and **3** clearly indicated the configuration at C-4. While **1** gave clear NOEs between H-14 and H-15 and H-6, the isomer **3** showed NOEs between H-14 and H-6 as well as between H-15 and H-5 and H-3. The configuration of **3** has been assigned previously for a lactone named barrelin [7]. Comparison of the ¹³C NMR data, however, show that this lactone most likely is identical with vulgarin though the mp and the optical rotation differ. The published ¹³C NMR data of **1** [8] differ only in the chemical shift of C-14 which was erroneously assigned (δ 22.7 is the value of C-8 and not of C-14 which is 19.7). The ¹³C NMR data of **1** and **3** show some clear differences. In particular, the C-15 signal is